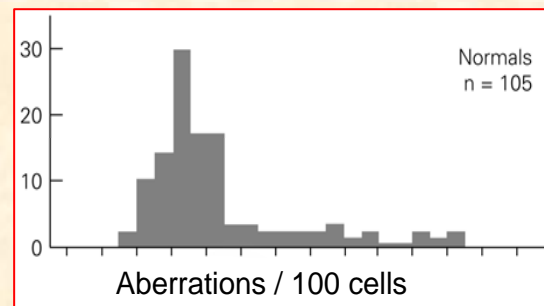


Possible High-Throughput Screening Logistics

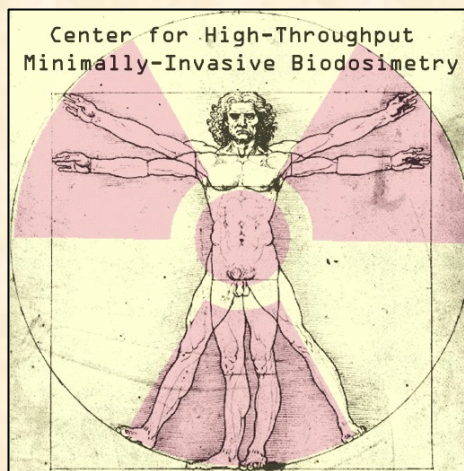
Integrating
*High-Throughput
Radiation Biodosimetry*
with
*High-Throughput Assays
for Radiation Sensitivity*



David J. Brenner
*Columbia Center for High-Throughput
Minimally-Invasive Radiation Biodosimetry*



Columbia Center for High-Throughput Minimally-Invasive Radiation Biodosimetry



www.cmc.columbia.edu



Ultra high-throughput biodosimetry

In response to a radiological event, small or large, in a major US city, tens or hundreds of thousands of people will need to be screened within a few days for radiation exposure....

- 1) for triage and treatment of acute radiation effects
- 2) for long term assessment of late effects (cancer, cardiac disease)
- 3) because active reassurance measures are an effective means of reducing mass panic

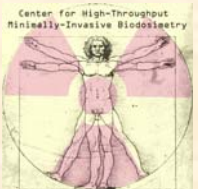
Current (manual) technologies allow screening of some tens of individuals per day

Issues for an Effective High-Throughput Biodosimeter

- ❖ Processing throughput – minimal invasiveness
- ❖ Sensitivity – dose coverage
- ❖ Specificity
- ❖ Processing time
- ❖ Signal stability

Need for more than one approach....

Different biodosimetric endpoints needed for different situations



Our High-Throughput Radiation Biodosimetry Approaches

PROGRAM 1: Converting current biomarkers to ultra-high throughput

Micronuclei and γ -H2AX: Both already well characterized. Amenable to automation; current systems have very limited throughputs.

PROGRAM 2: Genomically-based high-throughput biodosimetry

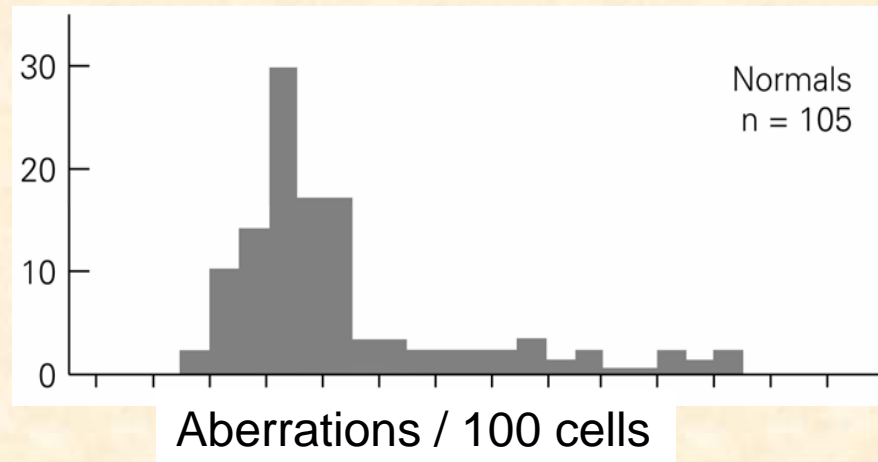
Gene expression profiling to provide a “signature” of radiation damage has been pioneered by Amundson and Fornace. The technology in a high-throughput context is new, but well advanced.

PROGRAM 3: Metabolomically-based high-throughput biodosimetry

Metabolomics (global metabolite profiles) has the potential to provide a rapid non-invasive radiation biodosimeter. The technology in a high-throughput context is well advanced.

High throughput biodosimetry is well established as critical for radiation threat countermeasures

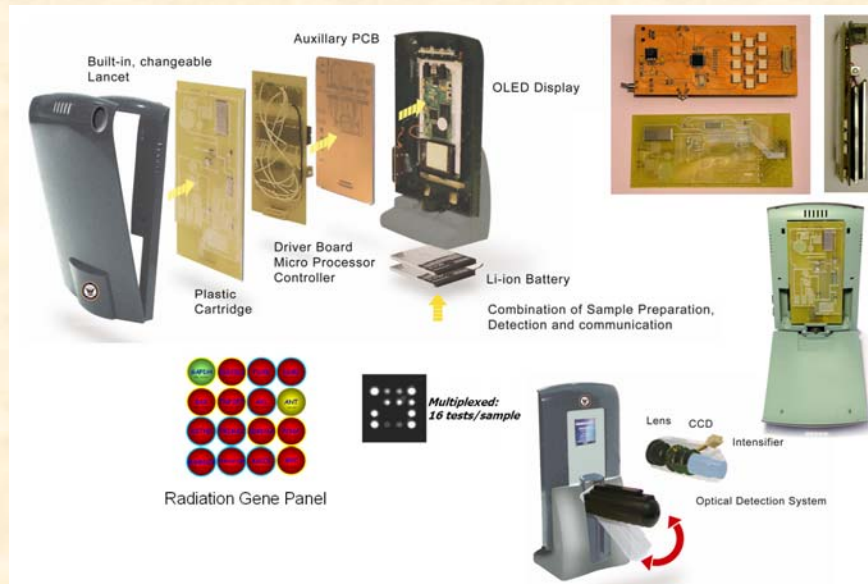
But less attention has been paid to high-throughput assays for radiation sensitivity....



....in large part because robust predictors of individual radiation sensitivity, the subject of this Workshop, have yet to be established.

In practice, how might a high-throughput assay of individual radiation sensitivity work?

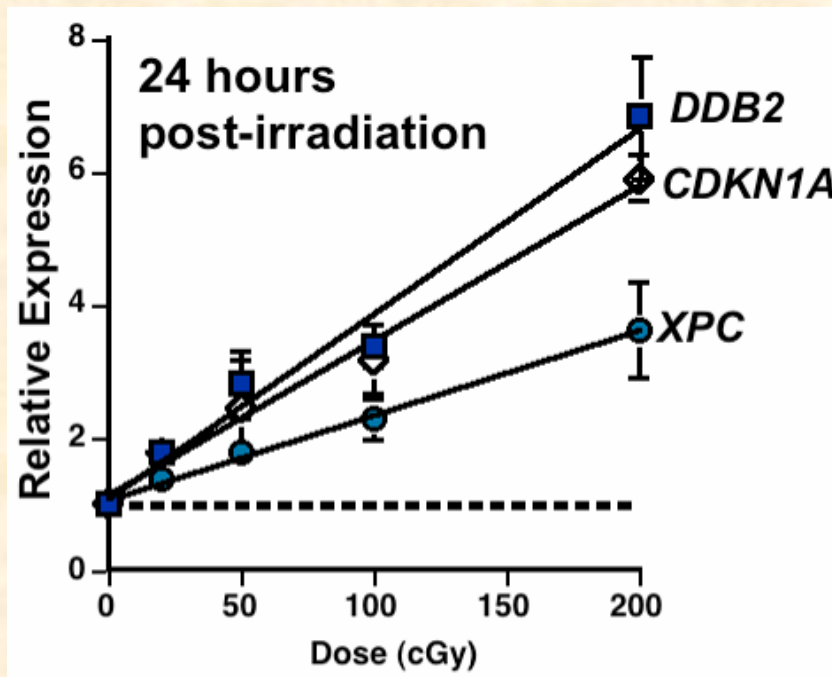
- ❖ **Could it be integrated with high-throughput biodosimetry?**



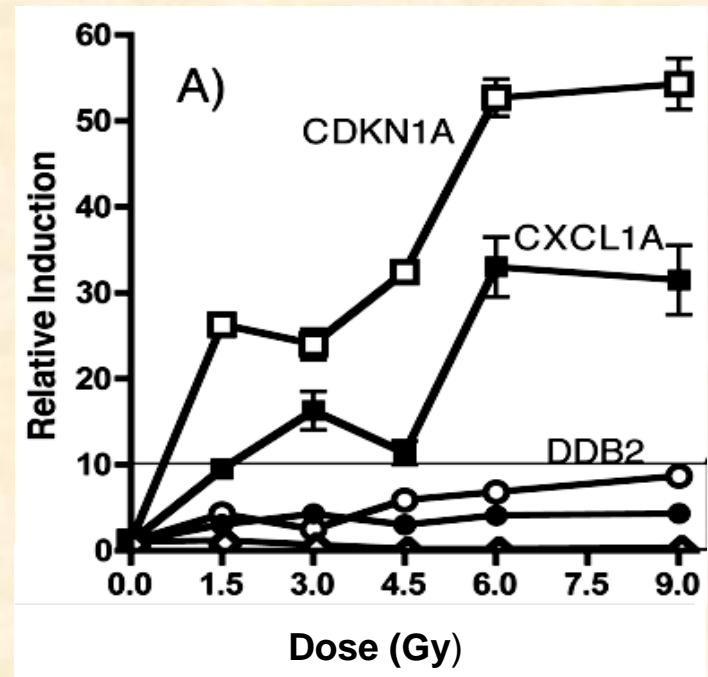
Gene expression as a basis for radiation biodosimetry

Exposure to ionizing radiation produces dose-dependent changes in the expression of many genes, potentially providing a means to assess both radiation exposure, and to quantify dose.

Ex vivo



Amundson *et al.*, (2000)
Radiation Research, 154 (3): 342-346



Amundson *et al.*, (2004)
Cancer Research, 64: 6368-6371

We have a US patent for this approach

The
United
States
of
America



UNITED STATES PATENT

Granted to

Sally A. Amundson, Albert J. Fornace, Jr.,
Jeffrey M. Trent

(12) United States Patent
Fornace, Jr. et al.

(10) Patent No.: US 7,008,768 B1
(45) Date of Patent: Mar. 7, 2006

(54) METHOD FOR DETECTING RADIATION
EXPOSURE

OTHER PUBLICATIONS

(75) Inventors: Albert J. Fornace, Jr., Bethesda, MD
(US); Sally A. Amundson, New York,
NY (US); Jeffrey M. Trent, Rockville,
MD (US)

(73) Assignee: The United States of America as
represented by the Department of
Health and Human Services,
Washington, DC (US)

(*) Notice: Subject to any disclaimer, the term of this
patent is extended or adjusted under 35
U.S.C. 154(b) by 0 days.

(21) Appl. No.: 08/913,171

(22) PCT Filed: Feb. 25, 2000

(86) PCT No.: PCT/US00/04897

\$ 371 (c)(3),
(1), (4) Date: Aug. 8, 2001

(87) PCT Pub. No.: WO00/20643

PCT Pub. Date: Aug. 31, 2000

Related U.S. Application Data

(40) Provisional application No. 60/121,756, filed on Feb.
26, 1999.

(51) Int. Cl.

C12Q 1/68 (2006.01)

C12P 1/24 (2006.01)

C07H 21/04 (2006.01)

(52) U.S. Cl.

435/6; 435/91.1; 435/91.2;
536/23.1

(58) Field of Classification Search 435/6;
435/91.1; 91.2; 536/23.1, 24.33
See application file for complete search history.

References Cited

U.S. PATENT DOCUMENTS

5,563,066 A 10/1996 Rader
5,690,157 A 10/1997 Gung et al.
5,707,801 A 1/1998 Sato
5,741,308 A 4/1998 Foster et al.
5,773,722 A 6/1998 Fockler et al.
5,830,042 A 10/1998 Plakol et al.
5,843,655 A 10/1998 Kottel
5,866,132 A 12/1998 Kottel et al.
5,877,212 A 8/1999 Richter et al.
5,913,300 A 6/1999 Ullrich
5,968,352 A 10/1999 Shugart et al.
5,998,136 A 12/1999 Kottel

FOREIGN PATENT DOCUMENTS

WO 90/01419 8/1997
WO 90/01860 5/1996
WO 90/20954 12/1996
WO 90/25913 11/1996

Amundson et al., "Identification of Gene-Ray Responses
Using cDNA Array Hybridization," *Proceedings of the
American Association for Cancer Research Annual Meeting*
35:454 (1996).

Amundson et al., "Induction of Stress Genes by Low Doses
of Gamma Rays," *Radiat. Res.* 152:225-231 (1999).

Amundson et al., "Fluorescent cDNA Microarray
Hybridization Reveals Complexity and Heterogeneity of
Cellular Genotoxic Stress Responses," *Oncogene* 18:3666-
3673 (1999).

Cacciari et al., "High-Throughput Analysis of Differential
Gene Expression," *J. Cell. Biochem. Suppl.* 30/31:286-296
(1998).

Daniel et al., "Use of a cDNA Microarray to Analyze Gene
Expression Patterns in Human Cancer," *Nature Genetics*
14:457-460 (1996).

Pisecchia et al., "Wip1, A Novel Human Protein Phosphatase
that is Induced in Response to Ionizing Radiation, in a
p53-Dependent Manner," *Proc. Natl. Acad. Sci. USA* 94:
6048-6053 (1997).

Fornace et al., "The Complexity of Radiation Stress
Responses: Analysis by Library and Fractional Genom-
ics Approaches," *Gene Expr.* 7:387-400 (1999).

Goley et al., "Induction of Serum Amyloid A Inflammatory
Response Genes in Irradiated Bone Marrow Cells," *Radiat.
Res.* 145:573-578 (1996).

Higuchi et al., "Search for Genes Involved in UV-Resistance
in Human Cells by mRNA Differential Display: Increased
Transcriptional Expression of Nucleophosmin and L-Fabrin
Genes in Association with the Necrosis," *Radiation
Environ. Res. Comm.* 248:577-602 (1998).

(Continued)

Primary Examiner—Kermit R. Horlitz,
Assistant Examiner—Young I. Kim
(74) Attorney, Agent, or Firm—Kleinfelder Spitznagel LLP

ABSTRACT

A method is disclosed for detecting exposure of organisms
to biologically significant or hazardous amounts of ionizing
radiation. The method uses nucleic acid microarray hybrid-
ization to evaluate biological effects, such as patterns of
expression of genes after radiation exposure. Nucleic acid
genes are provided which have been found to be responsive
to radiation exposure in a variety of cell lines. These genes
are incorporated into probe sets, which are exposed to a
labeled nucleic acid composition from a test cell, such as
cDNA or total RNA extracted from the test cell, which
specifically hybridizes to members of the probe set when the
cell has been exposed to a biologically significant amount of
ionizing radiation. Whether the nucleic acid composition
hybridizes to the nucleic acid molecules representing genes
that are differentially expressed is determined. The invention
also includes methods for determining a dose response
relationship between radiation exposure and differential
expression of one or more genes, the example to determine
a probable radiation dose in cells that have actually or
potentially been exposed to the ionizing radiation. The
invention also includes probe sets and microarrays used in
this method.

58 Claims, 11 Drawing Sheets

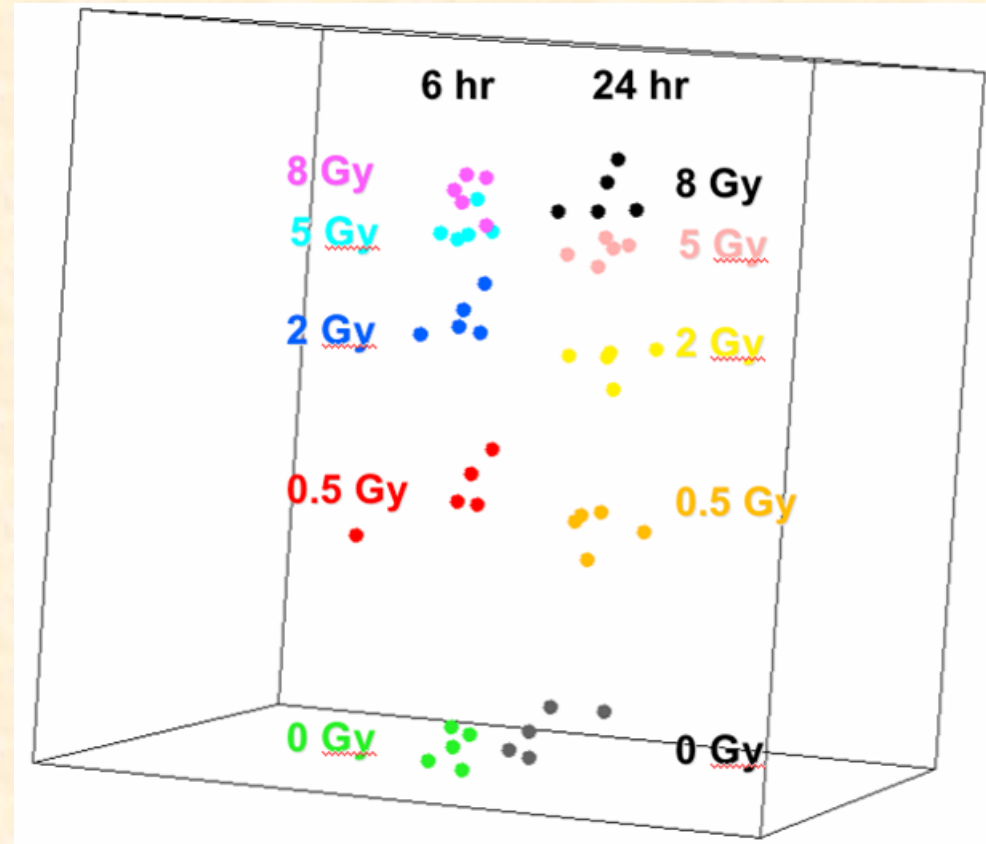
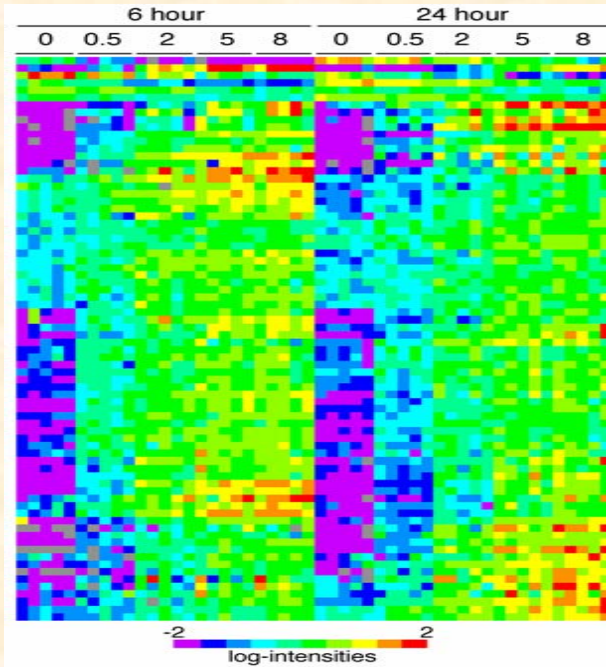
**"METHOD FOR DETECTING
RADIATION EXPOSURE"**

**US Patent # 7,008,768
March 7, 2006**

Center for High-Throughput
Minimally-Invasive Biodesign

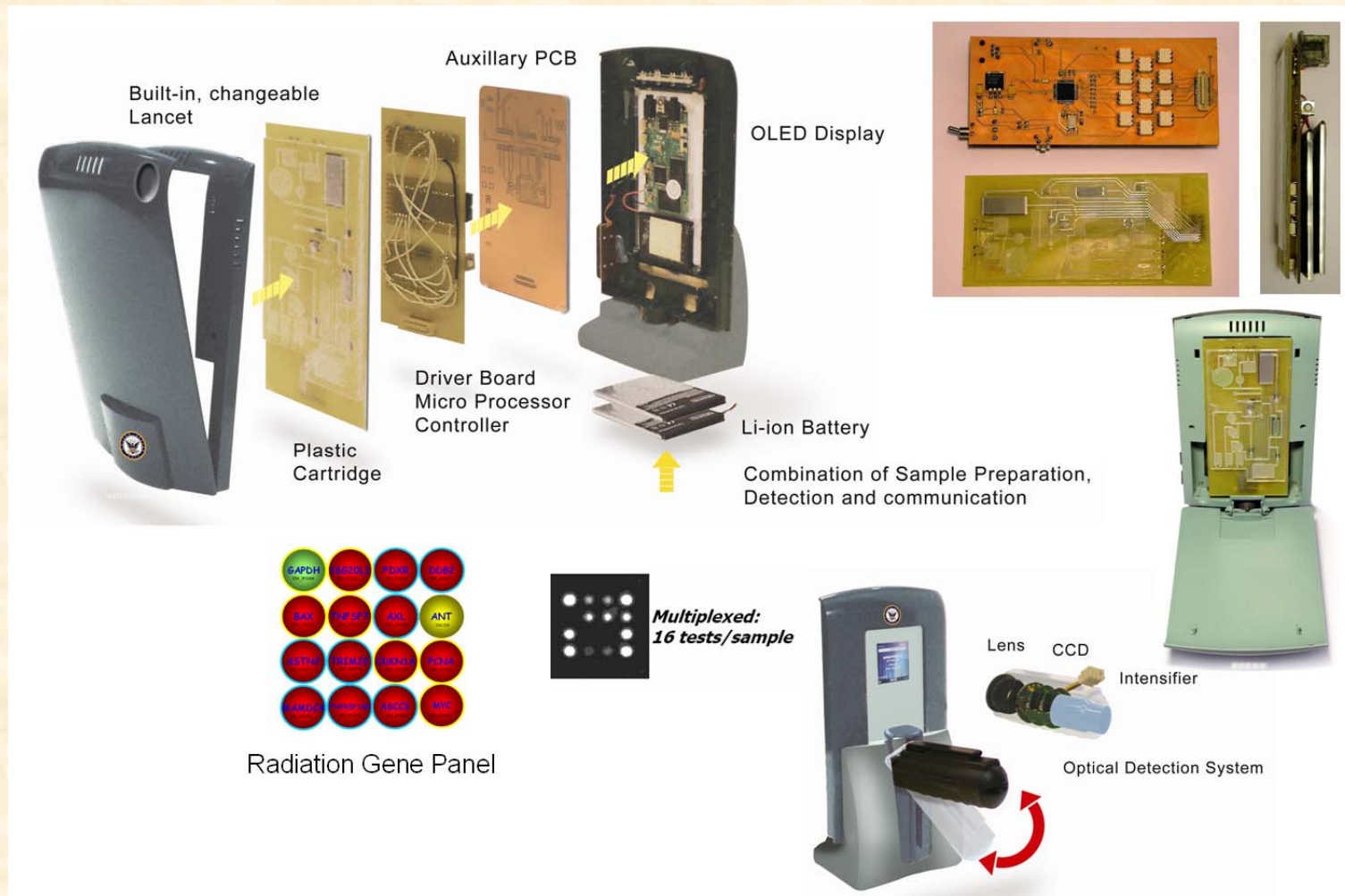


Multi-dimensional scaling (MDS) plot of a 74-gene response profile, separating samples by dose

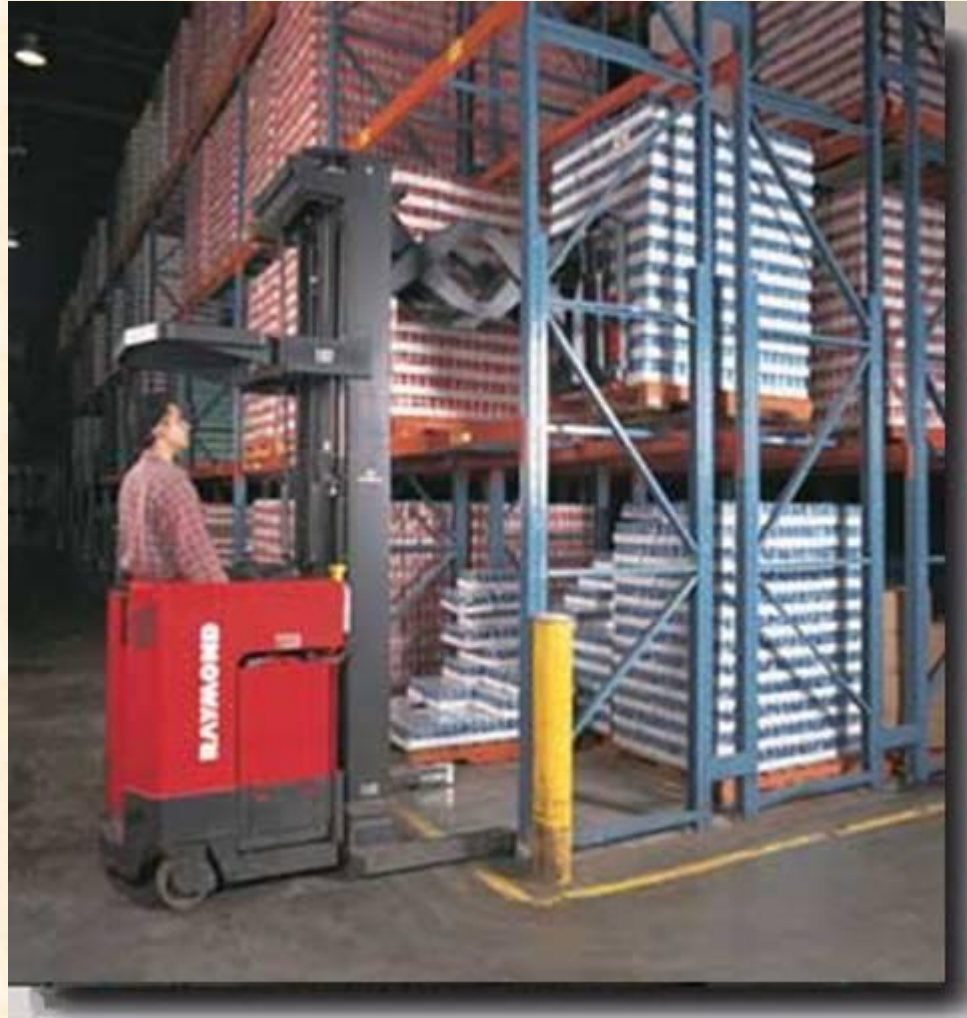


Using just a few genes does not give us the specificity we need

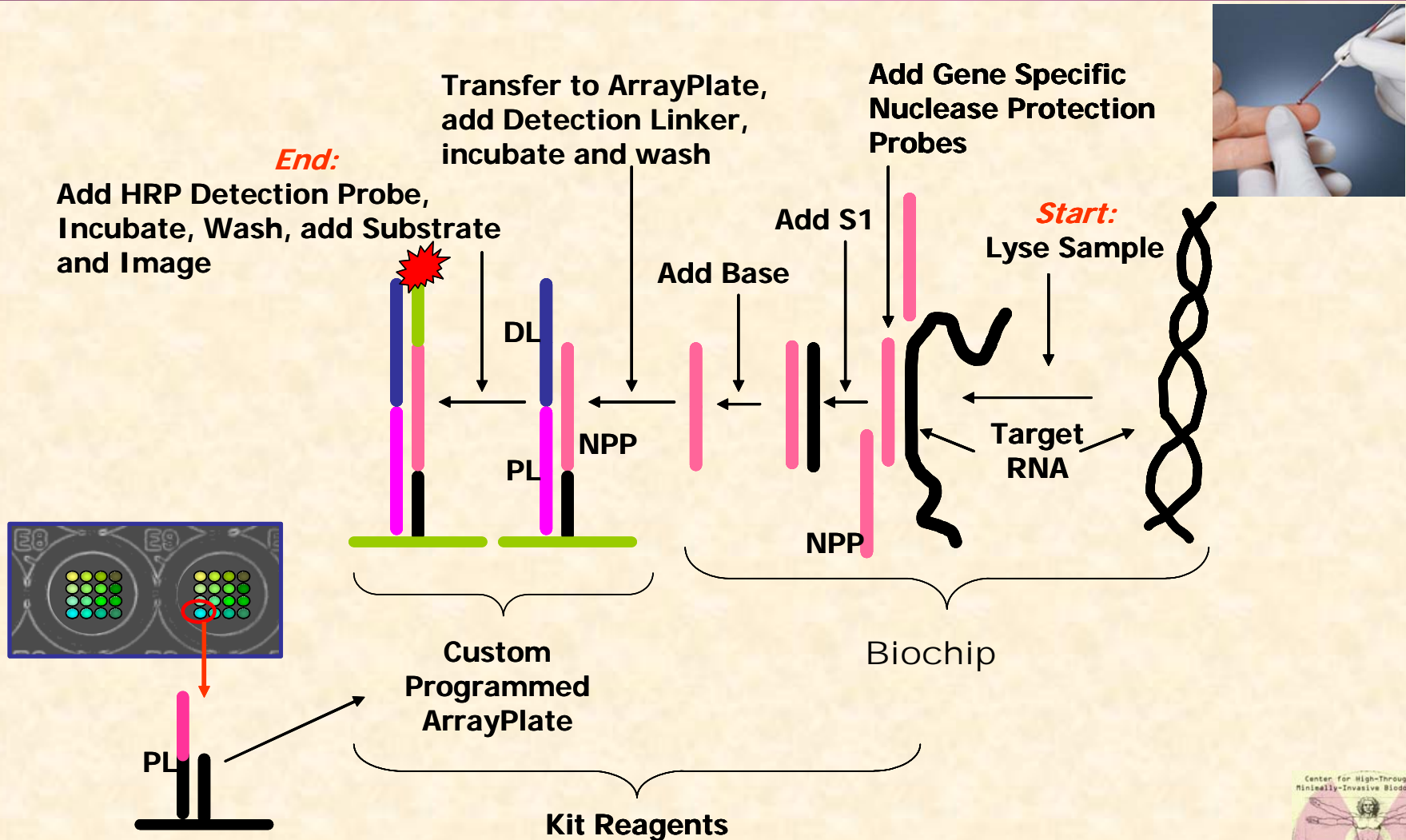
Components of genomically-based high throughput biodosimetry system



Thousands of such cartridges can be stockpiled,
for use after a large-scale radiological event



Quantitative Nuclease Protection Assay (qNPA) for High Throughput Genomics

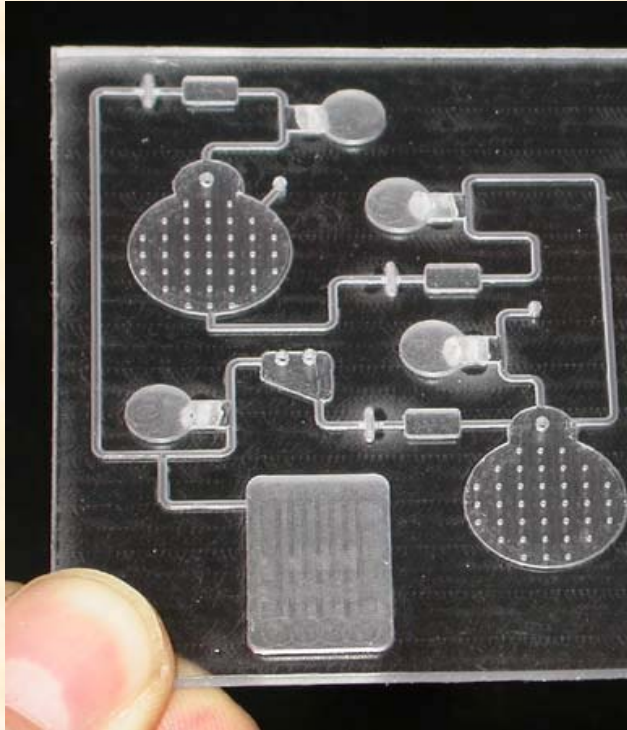


Advantages of qNPA (quantitative Nuclease Protection Assay)

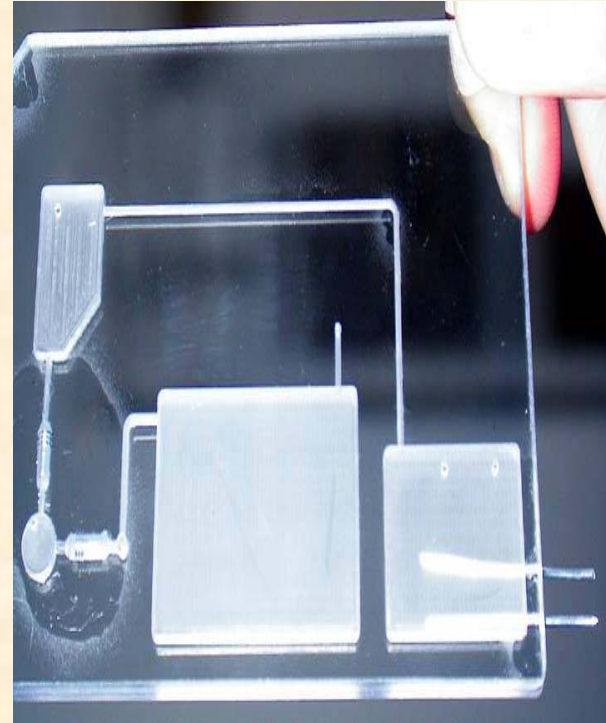
- ✓ **Whole blood is used**
Mix with lysis buffer, run assay.
- ✓ **Directly measures gene abundance:**
No amplifications (eg PCR) needed.
- ✓ **Multiplexed:**
Array simultaneously measures multiple genes
- ✓ **Reproducible and repeatable day-to-day:**
Variability between samples $\leq 10\%$, repeatability day-to-day within 2%.
- ✓ **Programmable:**
Change test genomic signature by use of a different set of reagents with the same microfluidic disposable array.



Sample analysis cartridge for qNPA



The *front-half* of the qNPA assay takes the place of the Nuclease Protection Plate of the standard microwell format



The *back half* of the qNPA assay takes the place of the Array Plate of the standard microwell format

Standalone DNA prototype



**Self-contained
battery operated
cassette for analysis
of DNA**

**Readily adaptable for
field assessment of
SNPs predicting
radiosensitivity**

Genomically-based identification of radiation-sensitive cancer cell lines

January 15, 2008
Volume 68
Number 2
Pages 339-626



Gene Module
Maps Target
Cancer Therapy
Page 369

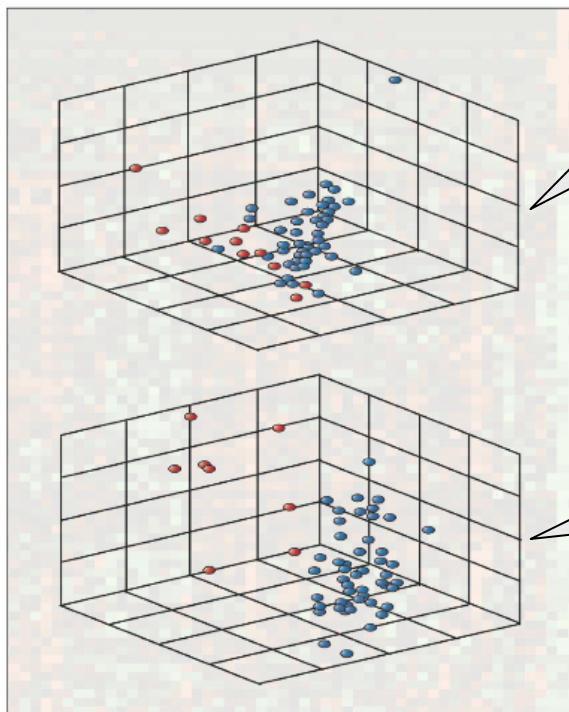


Soluble VEGFR-2
Levels as a
Surrogate Biomarker
for Tumor Growth
Page 521

Integrating
Global Gene
Expression and
Radiation Survival
Parameters
across the NCI
Drug Screen
Page 415



Cancer Research



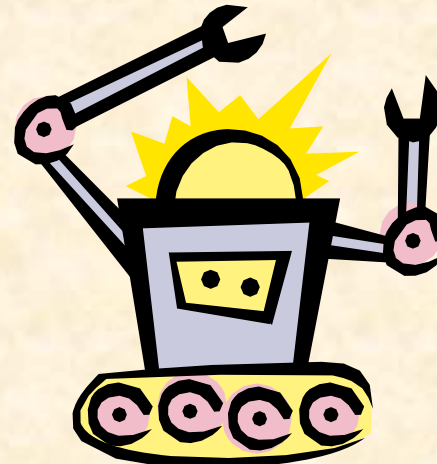
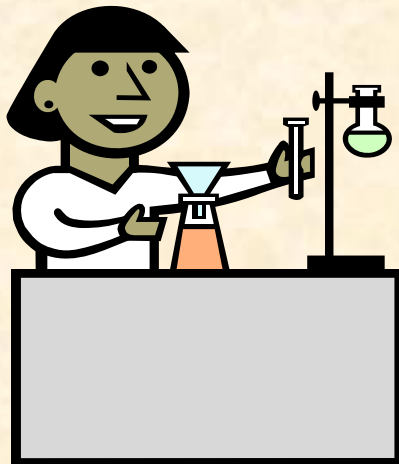
Radiation sensitive cell lines
(SF2 <0.2, red points)
separated by radiation-induced
expression of 22 genes

Radiation sensitive cell lines
(SF2 <0.2, red points)
separated by baseline
expression of 175 genes

RABIT:

Rapid Automated BIodosimetry Tool

Converting two manually-based radiation biodosimetry assays to high throughput, using a robotically-based biodosimetry workstation



RABIT: **R**apid **A**utomated **BI**odosimetry **T**ool

- Fully automated ultra high-speed robotic biodosimetry workstation.
- Automates two well-established manual assays, γ -H2AX and micronucleus
- One fingerstick of blood
- No human intervention



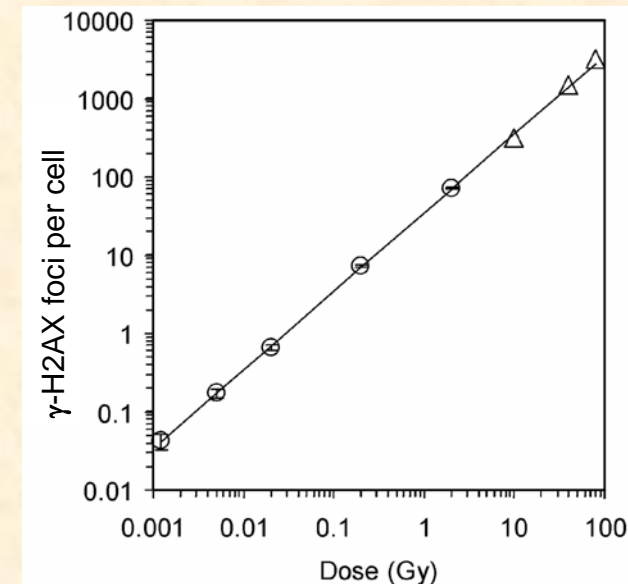
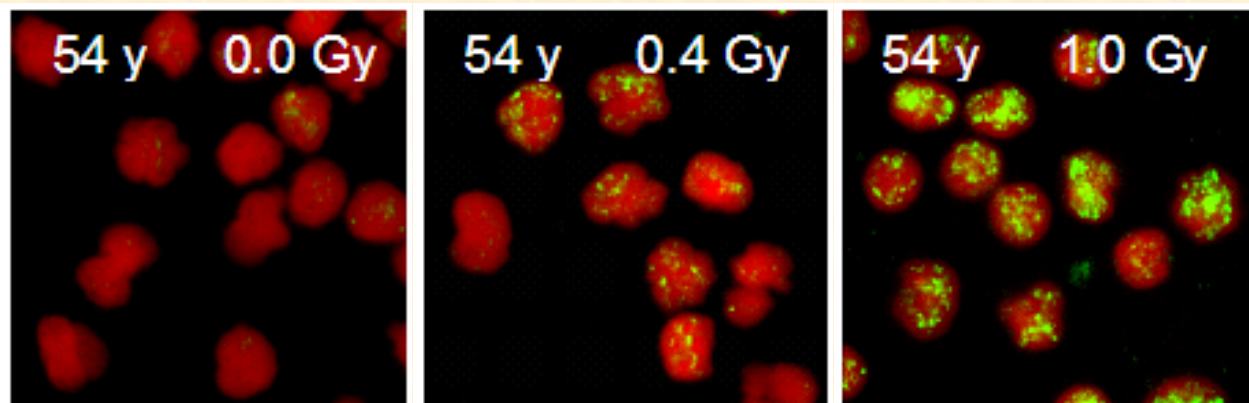
- **Phase I (2008):**
6,000 samples/day
- **Phase II (2010):**
30,000 samples/day

The main technical innovations are:

- 1) Use of smaller samples – single drop of blood from a capillary finger stick
- 2) Complete automation of biology and imaging in multi-well plates
- 3) Innovations in high-speed imaging

The γ -H2AX assay:

A validated biomarker of radiation dose



- Each green spot represents the location of a DNA repair complex
- Assay does not require culturing cells to mitosis

- Highly linear with radiation dose

Micronuclei: A validated biomarker of radiation dose

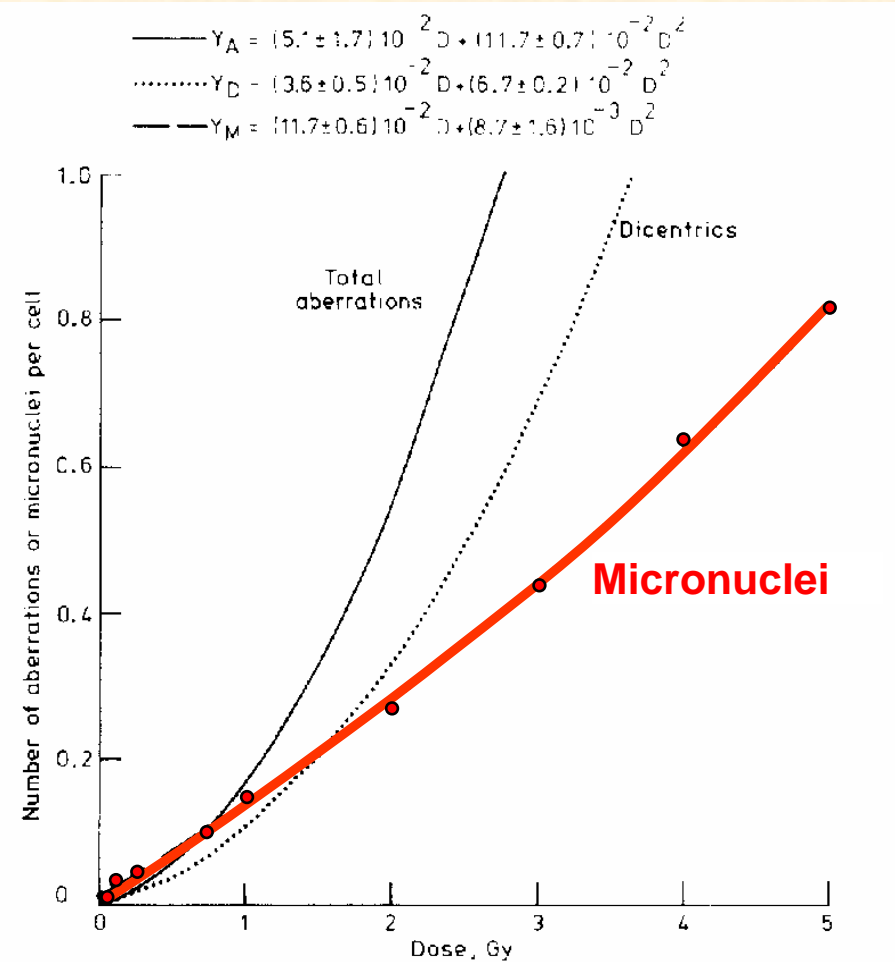


Fig. 1. Graph of micronuclei per cell against X-ray dose for all donors, fitted to the quadratic model, together with published dose-response curves for total aberrations and dicentric (Lloyd et al., 1986).

γ -H2AX vs. Micronuclei

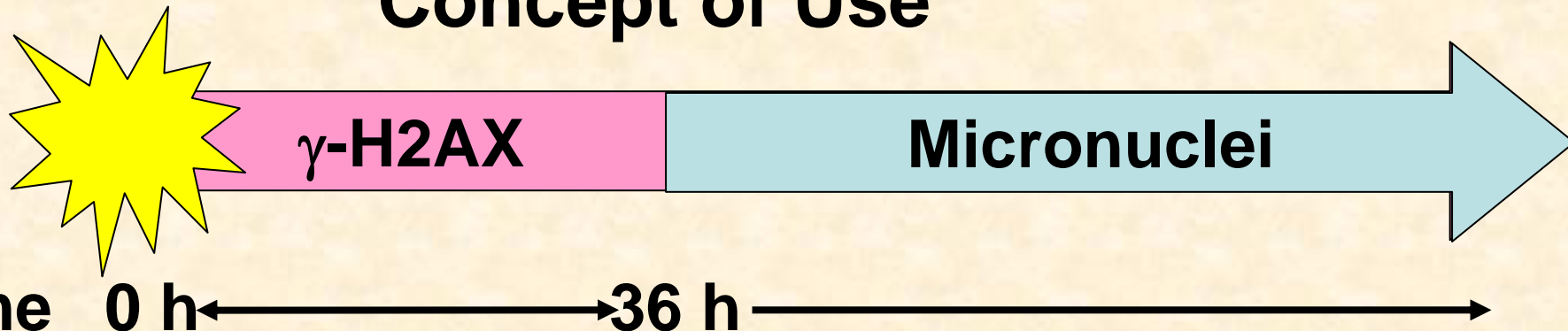
γ -H2AX

- 😊 Same day processing
- 😊 Highly linear with dose
- 😞 Signal lasts only ~36 h
- 😊 Amenable to high-throughput automation

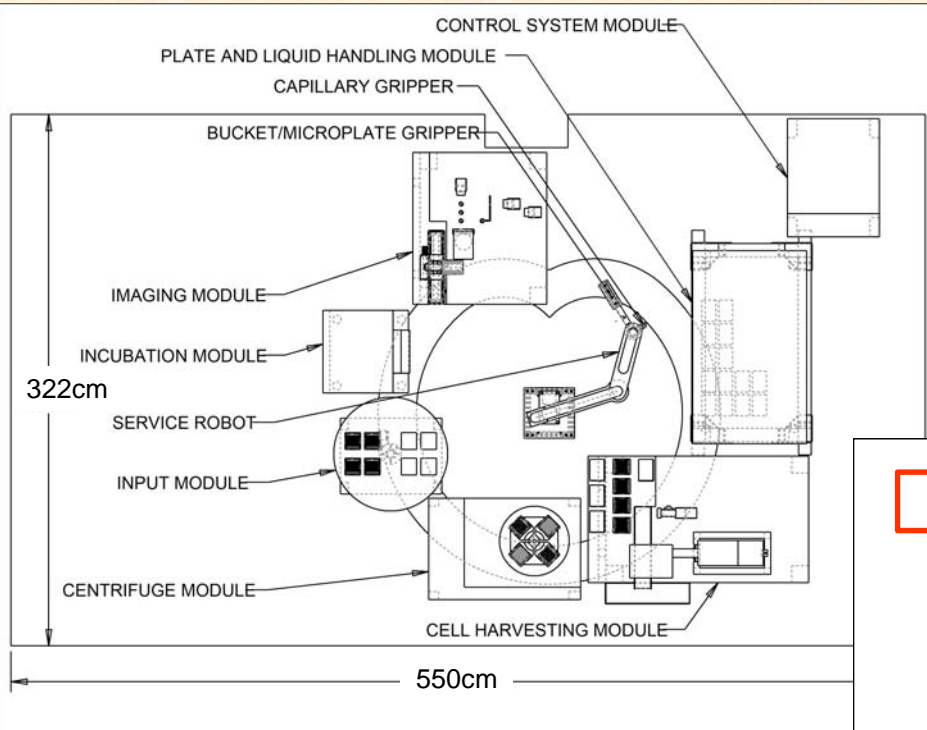
Micronuclei

- 😞 70 hour processing
- 😊 Slightly non linear with dose
- 😊 Signal stable for years
- 😊 Amenable to high-throughput automation

Concept of Use

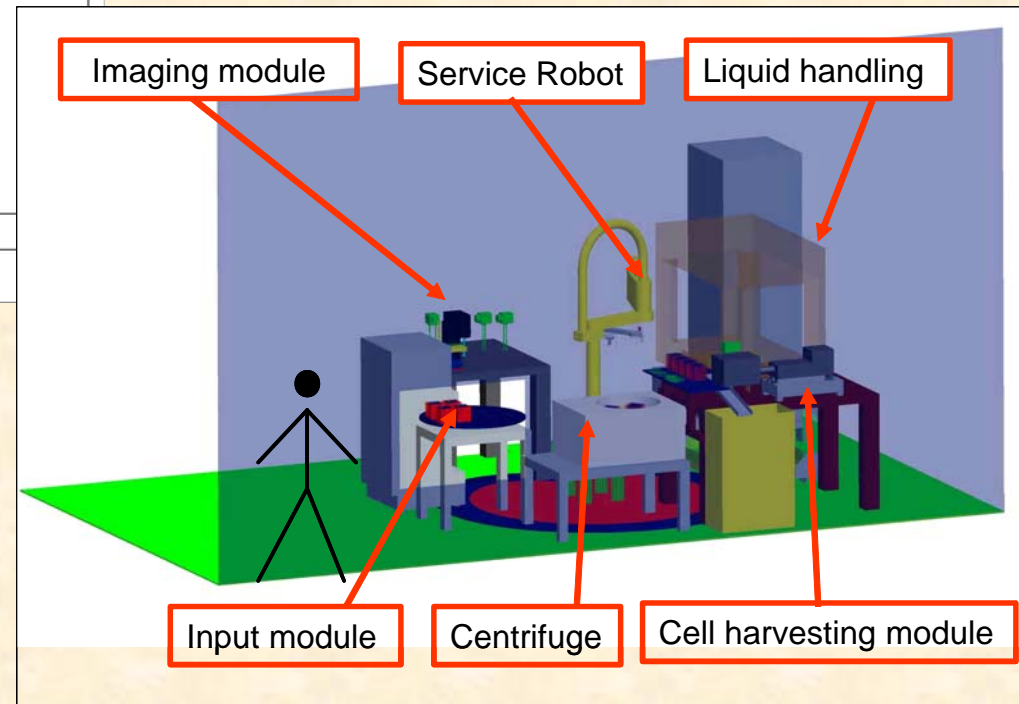


RABIT device overview

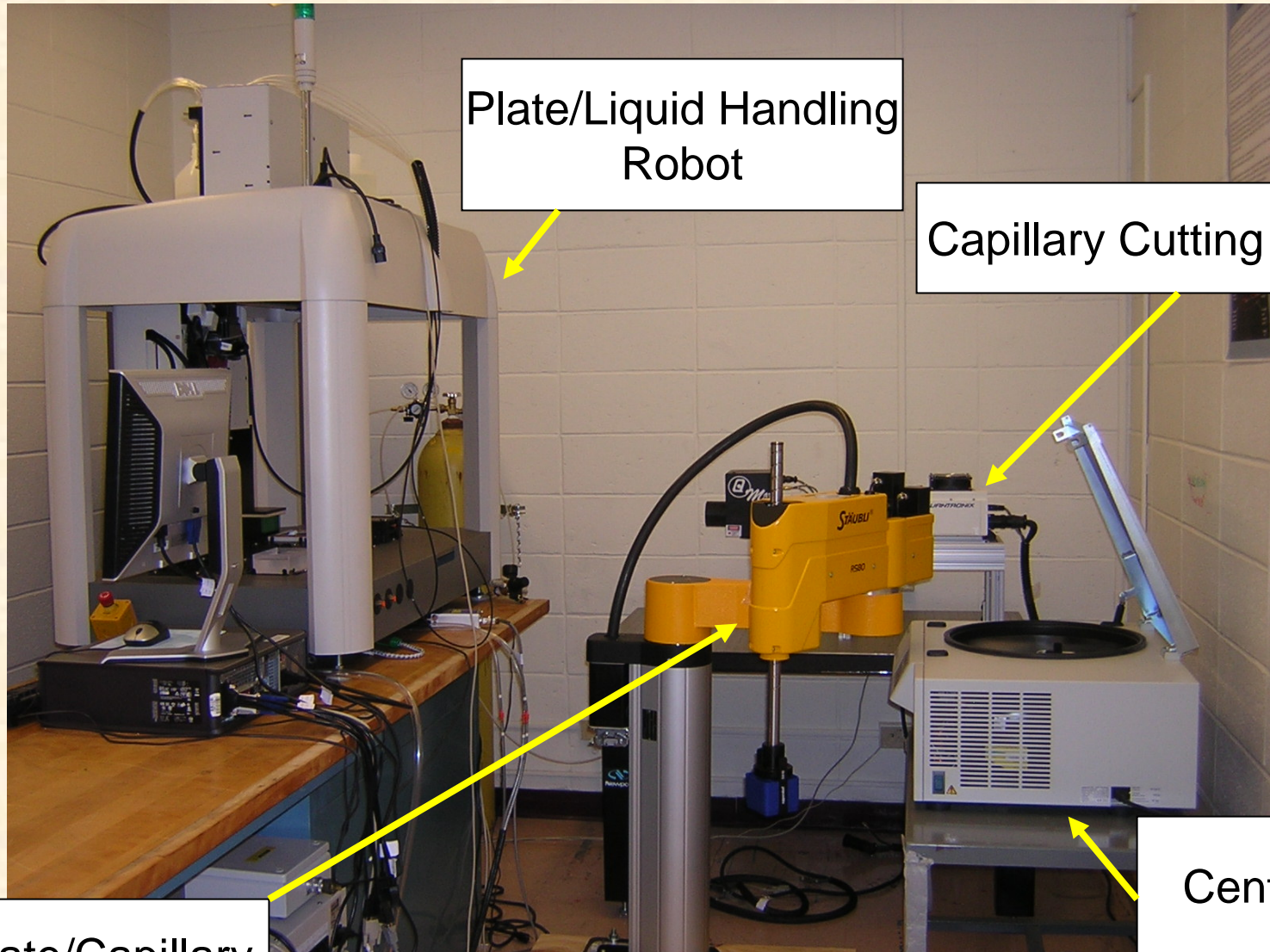


- **Modularity**
- **Truck portability
(3 x 5 m footprint)**

- **Full autonomy,
safe human access**
- **18 hour duty cycle
30,000 samples**



Breadboard Prototype



Plate/Liquid Handling
Robot

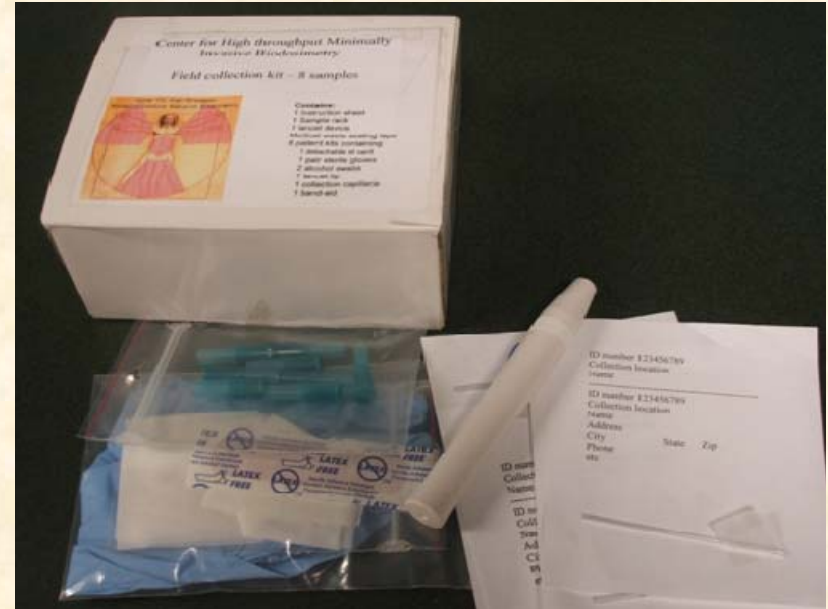
Capillary Cutting System

Centrifuge

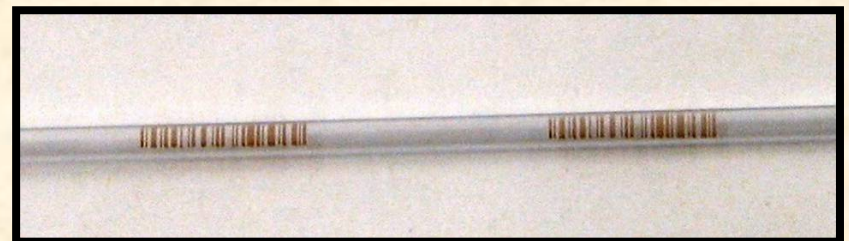
Bucket/Plate/Capillary
Handling Robot

RABIT Field Collection

- We anticipate multiple collection sites, at church halls, doctor's offices, etc.
- Standard capillary lancet used to draw drop of blood (same as used for home diabetes tests)
- When filled, capillary is placed in a 24-tube holder, designed for transport to (and direct insertion in) the **RABIT** machine



Field collection kit contains matched bar-coded capillaries, data collection cards, capillary holder, gloves, lancets, etc.





Central RABIT location



Using the RABIT to assess radiation sensitivity

- The RABIT would not be able to assay for a mechanistically-based predictor of radiosensitivity, but a more functional approach is an option.
- The sample is split into two (A and B):
 - A is assayed for the biodosimetric endpoint of choice;
 - B is irradiated, in a high throughput platform, to a dose significantly higher than those anticipated from the radiological event;
 - Then B is assayed in exactly the same way as A.
- Sample B will yield a functional estimate of radiation sensitivity, to augment the dose estimate from Sample A.



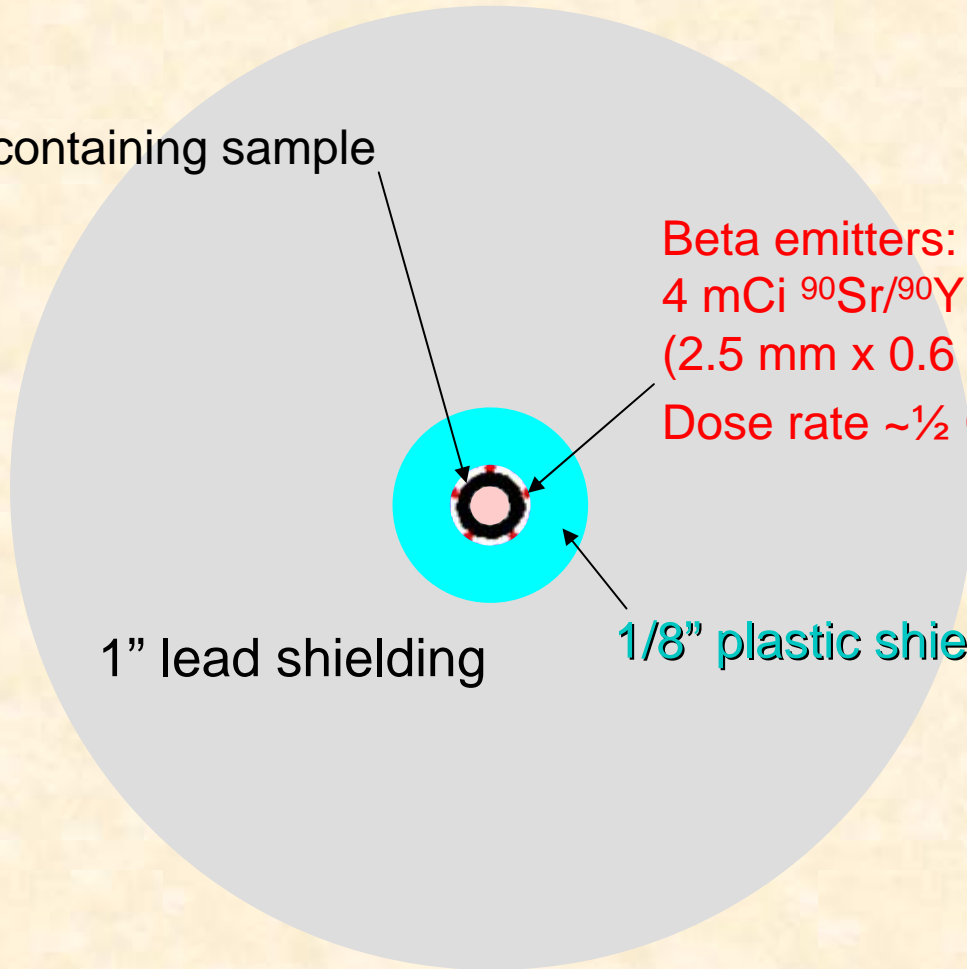
High throughput capillary irradiator

Capillary tube containing sample

Beta emitters:
4 mCi $^{90}\text{Sr}/^{90}\text{Y}$ seeds
(2.5 mm x 0.6 mm diam)
Dose rate $\sim \frac{1}{2}$ Gy / sec

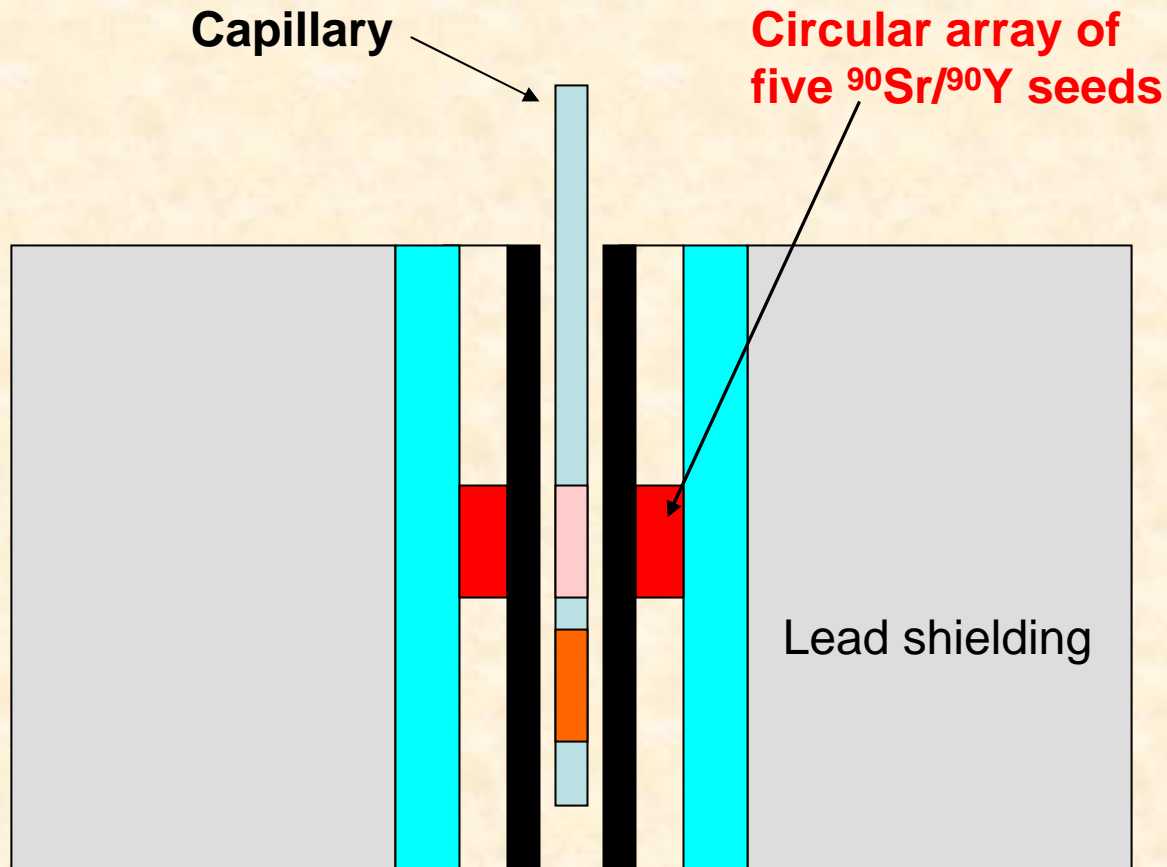
1" lead shielding

1/8" plastic shielding



Top view

High throughput capillary irradiator



Side view, not to scale

In practice, how might a high-throughput assay of individual radiation sensitivity work?

- ❖ **Could it be integrated with high-throughput biodosimetry?**

Yes

- ❖ **Mechanistically-based genomic radiosensitivity assays could indeed be integrated**
- ❖ **Functionally-based approaches are also feasible**